

NATURAL AND SYNTHETIC MATERIALS
WITH THE INSECT HORMONE ACTIVITY. XII.*

SYNTHESIS OF METHYL 3,7,11,11-TETRAMETHYL-2-DODECENOATE
AND SOME OF ITS HOMOLOGUES

M.ROMAŇUK, L.STREINZ and F.ŠORM

*Institute of Organic Chemistry and Biochemistry,
Czechoslovak Academy of Sciences, Prague 6*

Received April 2nd, 1971

Using Kolbe anodic synthesis several esters of aliphatic α,β -unsaturated C_{14} — C_{17} acids with a terminal quaternary C-alkyl group have been prepared.

In recent years ever greater attention is being devoted to substances imitating the effect of the insect juvenile hormone in the insect organism. Some of these substances were isolated from natural sources, but a much larger number of them were prepared synthetically¹⁻³. Both cecropia juvenile hormones *I*, *II* were isolated and identified⁴⁻⁶ some time ago, and some of their analogues were prepared synthetically^{1-3,7-10}, similarly to juvabione^{2,3,11} (*III*) and related aromatic derivatives with juvenilizing activity^{12,13}. Substances similar to farnesylic acid^{2,3,14-20} (*IV*) constitute another group of synthetic analogues.

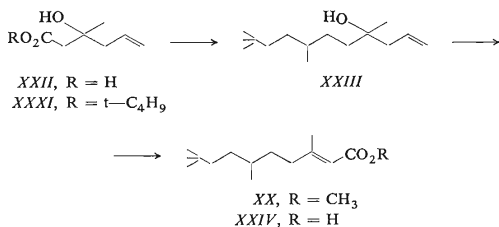
In our laboratories the selectivity of action of different analogues in dependence on structural changes in their molecule has been investigated.** In connection with the interesting physiological results obtained with methyl 11-chloro-3,7,11-trimethyl-2-dodecenoate (*V*) (ref.²⁰) we decided to synthesise its carbon analogues in which chlorine was substituted by a methyl group. This paper presents only chemical results. The relationship between the structure and activity has been discussed elsewhere²¹. As a key reaction we made use of Kolbe's anodic synthesis; this reaction was used for the preparation of some terpenic derivatives by Burrell and coworkers²². In principle the reaction represents the construction of the isoprenoid skeleton from several components; in each of them one of the tertiary alkyl substituted carbons is present (Scheme 1). For the synthesis of methyl 3,7,11,11-tetramethyl-2-dodecenoate (*VI*) we took 4,4-dimethylpentanoic acid (*VII*) (ref.²³) as the starting material which we obtained from the aldehyde *VIII* (ref.²⁴) by oxidation with silver oxide. Cross coupling of acid *VII* with monoester of β -methylglutaric acid (*IX*) (ref.²⁵)

* Part XI: *J. Insect Physiol.* 18, 19 (1972).

** In collaboration with the members of the Entomological Institute, Czechoslovak Academy of Sciences, Prague.

The use of 3,3-dimethylbutanoic acid (XV) (ref.²⁸) instead of homologue VII permitted us to prepare by the same reaction sequence shorter homologues of both esters VI and XIV. From acid XV and half-ester IX we thus obtained 3,6,6-trimethylheptanoic acid (XVI) which on Kolbe reaction with acid XI gave methyl 3,7,10,10-tetramethyl-2-undecenoate (XVII). In a similar manner 6,6-dimethyl-3-ethylheptanoic acid (XVIII), obtained from acid XV and monoester of β -ethylglutaric acid (XII), afforded in the presence of acid XI methyl 3,10,10-trimethyl-7-ethyl-2-undecenoate (XIX).

In connection with this work the preparation of methyl 3,6,9,9-tetramethyl-2-decenoate (XX) also became interesting, because it is, in actual fact, a bisnor derivative of ester VI. However, for its synthesis according to Scheme 1, the lower homolog of acid XI, *cis*, *trans*-3-methyl-4-methoxycarbonyl-3-butenoic acid (XXI) could not be used because the anodic synthesis with α,β - or β,γ -unsaturated acids does not take place in the expected manner²⁹. Therefore we employed a procedure represented in Scheme 2. By Kolbe reaction of 3,6,6-trimethylheptanoic acid (XVI) with 3-hydroxy-3-methyl-5-hexenoic acid (XXII) (ref.³⁰) we obtained 2,2,5,8-tetramethyl-10-undecen-8-ol (XXIII). Its ozonisation and dehydration gave a reaction mixture containing



SCHEME 2

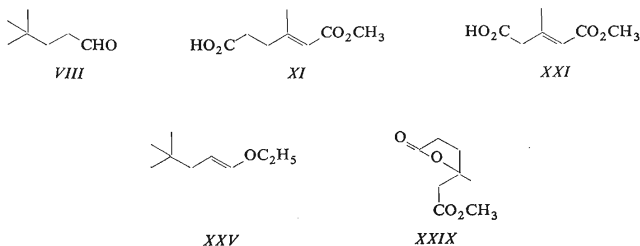


TABLE I
Anodic Synthesis

Product	Reaction components	B.p., °C/Torr	IR spectrum cm ⁻¹	n_D^{20} (t, °C)	Formula (mol.w.)	Calculated/Found	
						% C	% H
3,7,7-Trimethyloctanoic acid ^a (X)	VII	85-88	1 412, 1 710	1-4300	C ₁₁ H ₂₂ O ₂	70-92	11-90
	IX ^c	0-5	2 400-3 400	(23)	(186-3)	70-70	12-01
7,7-Dimethyl-3-ethyloctanoic acid ^a (XIII)	VII	89-93	—	1-4301	C ₁₂ H ₂₄ O ₂	71-95	12-08
	XIII ^d	0-2	—	(22)	(200-3)	72-23	12-21
3,6,6-Trimethylheptanoic acid ^a (XVI)	IX ^c	86-90 ^g	1 412, 1 709	1-4304	C ₁₀ H ₂₀ O ₂	69-72	11-70
	XV ^e	1-0	2 400-3 400	(24)	(172-3)	69-46	11-53
6,6-Dimethyl-3-ethylheptanoic acid ^a (XVIII)	XVII ^d	85-88	1 410, 1 710	1-4300	C ₁₁ H ₂₂ O ₂	—	—
	XV ^e	0-3	2 400-3 400	(23)	(186-3)	—	—
Methyl 3,7,11,11-Tetramethyl-2-dodecenoate ^b (VI)	X	110	1 154, 1 226	1-4533	C ₁₇ H ₃₂ O ₂	76-06	12-02
	XI ^f	0-3	1 366, 1 395 1 438, 1 650, 1 723	(22)	(268-4)	75-75	11-99
Methyl 3,11,11-Trimethyl-7-ethyl-2-dodecenoate ^b (XIV)	XI ^f	106-108	1 153, 1 227	1-4557	C ₁₈ H ₃₄ O ₂	76-54	12-13
	XIII	0-4	1 365, 1 395 1 438, 1 650, 1 723	(23)	(282-5)	76-37	12-01
Methyl 3,7,10,10-Tetramethyl-2-undecenoate ^b (XVII)	XI ^f	135-137	1 152, 1 367	1-4529	C ₁₆ H ₃₀ O ₂	75-53	11-89
	XVI	10	1 396, 1 650 1 721	(23)	(254-4)	75-57	11-83
Methyl 3,10,10-Trimethyl-7-ethyl-2-undecenoate ^b (XIX)	XI ^f	113-115	1 225, 1 366	1-4530	C ₁₇ H ₃₂ O ₂	76-06	12-02
	XVIII	0-5	1 390, 1 435 1 650, 1 720	(23)	(268-4)	76-26	12-05

^a Prepared by procedure A; ^b prepared by procedure B; ^c see ref.²⁵; ^d see ref.²⁷; ^e commercial preparation from Koch and Light; ^f see ref.²⁶; ^g m.p. 24-25°C.

in addition to other products 3,6,9,9-tetramethyl-2-decenoic acid (XXIV) and its methyl ester XX. The pure ester XX was obtained by esterification of this mixture with diazomethane and preparative chromatography on silica gel.

EXPERIMENTAL

Analytical thin-layer chromatography was carried out on silica gel (Merck, Kieselgel G nach Stahl), using a spray of phosphomolybdic acid in ethanol for detection. For column chromatography silica gel according to Pitra³¹ was used. For Kolbe anodic synthesis an apparatus was used consisting of a reaction vessel, cooling spiral, platinum electrodes according to Dolejš and Novotný³², a magnetic stirrer, and a thermometer. The course of the electrolysis was appreciably accelerated by stirring the reaction mixture during electrolysis, as well as by increased voltage of the direct current. The same effect was achieved by the occasional switching over of the electrodes. Such an acceleration of the electrolysis was necessary especially during work with unsaturated acids XI and XXII.

1-Ethoxy-4,4-dimethyl-1-pentene (XXV)

This²⁴ was prepared on reaction of tert-butylmagnesium chloride (from 74.8 g of tert-butyl chloride and 19.2 g of magnesium) with acrolein diethyl acetal (XXVI; 48.8 g). Fractional distillation through a column of 15 TP gave 23.7 g of product XXV, b.p. 75–78°C/55 Torr. For C₉H₁₈O (142.2) calculated: 75.99% C, 12.76% H; found: 76.21% C, 12.75% H.

4,4-Dimethylpentanoic Acid (VII)

A mixture of ether XXV and 10% sulfuric acid (200 ml) was refluxed for 1 hour. The formed 4,4-dimethylpentanal (VIII) was distilled off from the reaction mixture together with 100 ml of water into a flask in which it was immediately oxidised with silver oxide prepared from 10.4 g of silver nitrate and 6.4 g of potassium hydroxide. The reaction mixture was worked up in the conventional manner. The whole procedure was repeated four times. Totally 11.28 g of acid VII were obtained, b.p. 75–78°C/12 Torr, n_D^{20} 1.4159. For C₇H₁₄O₂ (130.2) calculated: 64.58% C, 10.84% H; found: 64.86% C, 11.01% H.

Anodic Syntheses

Procedure A: A solution of methyl hydrogen β -alkylglutarate (25 mmol) and the monocarboxylic acid (40 mmol) in methanol (30 ml) in which 50 mg of sodium were dissolved was electrolysed at 6 V and 30–35°C. The end of the electrolysis was indicated by the change of the pH value of the reaction mixture to higher values. After the evaporation of methanol the sample was taken into ether and chromatographed on a 100 fold amount of silica gel. Elution with a mixture of light petroleum-ether 9 : 1 gave a methyl ester which after 2 hours saponification with aqueous ethanolic potassium hydroxide afforded the corresponding saturated monocarboxylic acid (Table I).

Procedure B: Equimolar mixture of the monocarboxylic acid obtained by procedure A and *cis*, *trans*-4-methyl-5-methoxycarbonyl-4-pentenoic acid (XI) was electrolysed in methanol at 24 V and 35–45°C. After electrolysis the excess methanol was evaporated and the product purified by column chromatography on a 100 fold amount of silica gel with light petroleum-ether mixture (5 : 1) (Table I).

3-Hydroxy-3-methyl-5-hexenoic Acid (XXII)

Using tert-butyl acetoacetate (26.3 g), allyl bromide (26.5 g), and zinc shavings (14.9 g) tert-butyl 3-hydroxy-3-methyl-5-hexenoate (XXXI; 23.01 g) was obtained which on saponification with

aqueous alcoholic potassium hydroxide gave hydroxy acid *XXII* (ref.³⁰) (14.2 g), b.p. 94°C/1 Torr and n_D^{24} 1.4625. For $C_7H_{12}O_3$ (144.2) calculated: 58.31% C, 8.39% H; found: 58.33% C, 8.46% H.

2,2,5,8-Tetramethyl-10-undecen-8-ol (*XXIII*)

Anodic synthesis was carried out by a procedure similar to the preceding cases from 3,6,6-trimethylheptanoic acid (*XVI*; 3.50 g) and 3-hydroxy-3-methyl-5-hexenoic acid (*XXII*; 3.50 g). Chromatography on a 100 fold amount of silica gel with light petroleum-ether 7 : 1 and distillation gave 0.82 g of substance *XXIII*, b.p. 87–90°C/0.5 Torr and n_D^{23} 1.4500. The substance had in its IR spectrum characteristic maxima at 918, 1000, 1367, 1393, 1639, 3005, 3070, and 3610 cm^{-1} . For $C_{15}H_{30}O$ (226.4) calculated: 79.57% C, 13.36% H; found: 79.76% C, 13.49% H.

Methyl 3,6,9,9-Tetramethyl-2-decenoate (*XX*)

Alcohol *XXIII* (400 mg) was ozonised at –20°C in ethyl acetate (15 ml) until the reaction mixture turned blue. It was acidified with acetic acid (50%; 30 ml) and additioned with hydrogen peroxide (30%; 3 ml) and refluxed for one hour. After cooling and dilution with water (20 ml) the reaction mixture was extracted with ether (totally 100 ml). The combined ethereal fractions were dried over sodium sulfate and esterified with diazomethane. The crude dry residue (310 mg) was heated with formic acid (98%; 4.0 g) at 80–85°C for 2 hours and then allowed to stand overnight at room temperature. After evaporation of the major part of formic acid *in vacuo* the residue was esterified with ethereal diazomethane solution. Ether was evaporated and the residue chromatographed on a silica gel column (50 g). Elution with a light petroleum-ether mixture (4 : 1) and subsequent distillation gave 46 mg of ester *XX*, b.p. 90°C/1 Torr. Principal bands in the IR spectrum occurred at 1370, 1395, 1655, and 1715 cm^{-1} . Mass spectrum: molecular ion $M^+ = 240$ m/e , further 184 m/e ($M^+ - C_4H_8$), 209 m/e ($M^+ - CH_3O$), 59 m/e ($COOCH_3$), and 57 m/e (C_4H_9).

Elemental analyses were carried out in our laboratories by Mrs V. Rusová, E. Šýkorová, and Mr V. Štěrba. Infrared spectra were measured by Mrs K. Matoušková on a Zeiss UR 10 apparatus. We thank Dr J. Smolíkova for help in their interpretation. The purity of compounds was controlled with a Perkin Elmer F 11 gas chromatograph provided with FID. The columns were filled with silicone elastomer E 301 (2.5% on AW DCMS Chromosorb G) and the analysis was performed by Dr V. Lukeš and Mrs S. Holubová. Mass spectra were measured by Dr L. Dolejš, and Mrs M. Vokáčová on a MS 902 mass spectrometer, and we also thank Dr L. Dolejš for their interpretation.

REFERENCES

1. Röller H., Dahm K. H.: *Recent Progress in Hormone Research*, Vol. 24, p. 651. Academic Press, New York 1968.
2. Berkoff C. E.: *Quart. Rev.* 23, 372 (1969).
3. Cizin J. S., Drabkina A. A.: *Uspechi Chim.* 39, 1074 (1970).
4. Meyer A. S., Schneiderman H. A., Hanzmann E., Ko J. H.: *Proc. Natl. Acad. Sci. US* 60, 853 (1968).
5. Röller H., Bjerke J. S., Holthaus L. M., Norgard D. W., McShan W. H.: *J. Insect Physiol.* 15, 379 (1969).
6. Meyer A. S., Hanzmann E., Schneiderman H. A., Gilbert L. I., Boyette M.: *Arch. Biochem. Biophys.* 137, 190 (1970).
7. Findlay J. A., MacKay W. D.: *Chem. Commun.* 1969, 733.
8. Van Tamelen E. E., McCormick J. P.: *J. Am. Chem. Soc.* 92, 737 (1970).

9. Loew P., Siddall J. B., Spain V. L., Werthemann L.: Proc. Natl. Acad. Sci. US 67, 1462 (1970).
10. Loew P., Sidall J. B., Spain V. L., Werthemann L.: Proc. Natl. Acad. Sci. US 67, 1824 (1970).
11. Pawson B. A., Cheung H. C., Gurbaxani S., Saucy G.: J. Am. Chem. Soc. 92, 336 (1970).
12. Bowers W. S.: Science 161, 895 (1968).
13. Bowers W. S.: Science 164, 323 (1966).
14. Bowers W. S., Thompson M. J., Uebel E. C.: Life Sci. 4, 2323 (1965).
15. Romaňuk M., Sláma K., Šorm F.: Proc. Natl. Acad. Sci. US 57, 349 (1966).
16. Sláma K., Romaňuk M., Šorm F.: Biol. Bull. 136, 91 (1969).
17. Jarolím V., Hejno K., Sehnal F., Šorm F.: Life Sci. 8, 831 (1969).
18. Wakabayashi N.: J. Medicinal Chem. 12, 191 (1969).
19. Wakabayashi N., Sonnet P. E., Law M. W.: J. Medicinal Chem. 12, 911 (1969).
20. Sláma K., Hejno K., Jarolím V., Šorm F.: Biol. Bull. 139, 222 (1970).
21. Sláma K., Romaňuk M., Šorm F.: J. Insect. Physiol. 18, 19 (1972).
22. Burrell J. W. K., Garwood R. F., Jackmann L. M., Oskay E., Weedon B. C. L.: J. Chem. Soc. 1966, 2144.
23. Brändström A.: Acta Chem. Scand. 13, 613 (1959).
24. Quelet R., Bercot P., d'Angelo J.: Bull. Soc. Chim. France 1966, 3258.
25. Ställberg-Stenhagen S.: Arkiv Kemi 26A, 1 (1948).
26. Linstead R. P., Lunt J. C., Weedon B. C. L., Shephard B. R.: J. Chem. Soc. 1952, 3621.
27. Cason J., Gastaldo C., Glusker D. L., Aelinger J., Ash L. B.: J. Org. Chem. 18, 1129 (1953).
28. Gryszkiewicz-Trochimowski E., Gryszkiewicz-Trochimowski O.: Bull. Soc. Chim. France 1951, 269.
29. Weedon B. C. L.: Quart. Rev. 6, 380 (1952).
30. Tschesche R., Machleidt M.: Ann. 631, 61 (1960).
31. Pitra J., Štěrba J.: Chem. listy 56, 544 (1962).
32. Dolejš L., Novotný L.: This Journal 19, 716 (1954).

Translated by Ž. Procházka.